Add-On 10 / Safety Information*



Safety Information for DNA Isolation Part 1 and Part 2

This Add-on kit is only to be used in combination with the following kits: $SepsiTest^{TM}$ -UMD or UMD- $SelectNA^{TM}$!

Important notes: When working with chemicals, always wear suitable protective lab clothing and work in a Class II biological safety cabinet.

CAUTION: Never add hypochlorite (bleach) or acidic solutions directly to the sample-preparation waste.

Buffer *CM* contains guanidine hydrochloride, which can form highly reactive compounds and toxic gases when combined with hypochlorite or other acidic solutions.

For more information, please consult the appropriate material safety data sheets (MSDS) which are available on request.

Continue with safety classification of the components on page 2

*Please note: Before using this scheme inform yourself of the details of the procedure. Special care is required for working under DNA-free conditions and secure working conditions. Please consult the manual for more information.

Version 03

Add-On 10 / Safety Information*



Safety Information for DNA Isolation Part 1 and Part 2

Component	Classification and Hazard / Precautionary Statements*

Buffer CM

Contains guanidine hydrochloride (>10%)

Acute toxicity (oral) and irritating (eyes and skin)

H302-H315-H319; P301+P312-P302+P352-P305+P351+P338



* H302: Harmful if swallowed;

H315: Causes skin irritation;

H319: Causes serious eye irritation;

P301+P312: IF SWALLOWED: Rinse mouth. Do NOT induce vomiting;

P302+P352: IF ON SKIN: Wash with plenty of soap and water;

P304+P340: IF INHALED: Remove to fresh air and keep at rest in a position comfortable for breathing;

P305+P351+P338: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

Emergency call: Poison Information Center Mainz, **Germany,** Tel.: +49-6131-19240 / 24h (German, English, French). <u>Outside of Germany:</u> Please contact the regional company representation in your country.

Add-On 10 / DNA Isolation* Part 1: Add-on kit for SepsiTestTM-UMD



Protocol 1:

Medium Size Sample DNA Isolation (>1 to 5ml Body Liquids)

To be used with **Kit 1** and **Kit 2** of **SepsiTest**TM**-UMD**:

Kit 1 and Add-On 10 Kit: Extraction Buffers (+18 to +25°C):

Unpack buffer bottles (*Add-on 10*) and buffer vials (Kit 1, package A; briefly centrifuge). Place bottles and vials in the following order:

Bottles (Add-On 10): SU - CM - DB1

Vials (Kit 1): RS - RL - RP - CS - AB - WB - WS - ES

Kit 2: Enzymes & Reagents (-15 to -25°C):

A) Fill up procedure for samples less than 5ml volume

Samples >1ml and less than 5ml are filled up using buffer *SU* (*Add-On 10*). Transfer the sample by pipetting into a *50ml Centrifuge Tube* (*Add-On 10*). Then add buffer *SU* using a disposable 5ml pipette or pipette tip until reaching the 5ml mark of the tube. Discard pipette/pipette tip with excess buffer *SU*. Continue with part B.

B) Sample pre-treatment and DNA Isolation

Per sample:

- 1. Pipette **5ml sample** or **filled-up sample** (see part A) into a *50ml Centrifuge Tube*.
 - Add **2ml buffer** *CM* (Kit 1, package A), vortex for 15s. Let stand at room temperature (+18 to +25°C) for 5min.
- Add 2ml buffer DB1 (Kit 1, package A).
 Add 10μl MolDNase B (Kit 2), vortex for 15s.
 Incubate at room temperature (+18 to +25°C) for 15min.
- 3. Centrifuge at 9,500xg, 10min. Carefully decant supernatant.
- 4. Resuspend pellet in **1ml buffer RS** (Kit 1, package A). Transfer by pipetting into a *ST tube* (Kit 1, package B).

Continue with Short Manual of SepsiTestTM-UMD (page 5, step 5).

Add-On 10 / DNA Isolation* Part 1: Add-on kit for SepsiTestTM-UMD



Protocol 2:

Large Size Sample DNA Isolation (>5 to 10ml Body Liquids)

To be used with **Kit 1** and **Kit 2** of **SepsiTest**TM**-UMD**:

Kit 1 and Add-On 10 Kit: Extraction Buffers (+18 to +25°C):

Unpack buffer bottles (*Add-on 10*) and buffer vials (Kit 1, package A; briefly centrifuge). Place bottles and vials in the following order:

Bottles (*Add-On 10*): *SU - CM - DB1*

Vials (Kit 1): RS - RL - RP - CS - AB - WB - WS - ES

Kit 2: Enzymes & Reagents (-15 to -25°C):

A) Fill up procedure for samples less than 10ml volume

Samples >5ml and less than 10ml are filled up using buffer *SU* (*Add-On 10*). Transfer the sample by pipetting into a *50ml Centrifuge Tube* (*Add-On 10*). Then add buffer *SU* using a disposable 5ml pipette or pipette tip until reaching the 10ml mark of the tube. Discard pipette/pipette tip with excess buffer *SU*. Continue with part B.

B) Sample pre-treatment and DNA Isolation

Per sample:

- Pipette 10ml sample or filled-up sample (see part A) into a 50ml Centrifuge Tube.
 - Add **4ml buffer** *CM* (Kit 1, package A), vortex for 15s. Let stand at room temperature (+18 to +25°C) for 5min.
- Add 4ml buffer DB1 (Kit 1, package A).
 Add 10µl MolDNase B (Kit 2), vortex for 15s.
 Incubate at room temperature (+18 to +25°C) for 15min.
- 3. Centrifuge at 9,500xg, 10min. Carefully decant supernatant.
- 4. Resuspend pellet in **1ml buffer RS** (Kit 1, package A). Transfer by pipetting into a *ST tube* (Kit 1, package B).

Continue with Short Manual of SepsiTestTM-UMD (page 5, step 5).

*Please note: Before using this scheme inform yourself of the details of the procedure. Special care is required for working under DNA-free conditions and secure working conditions. Please consult the manual for more information.

Version 03

Add-On 10 / DNA Isolation* Part 2: Add-on kit for *UMD-SelectNA*TM



Protocol 1:

Medium Size Sample DNA Isolation (>1 to 5ml Body Liquids)

To be used with **Kit 1** and **Kit 2** of **UMD-SelectNA**TM:

Kit 1 and Add-On 10 Kit: Extraction Buffers (+18 to +25°C):

Unpack buffer bottles: SU - CM - DB1 (Add-On 10) - RS - RL (Kit 1)

Kit 2: Enzymes & Reagents (-15 to -25°C):

A) Fill up procedure for samples less than 5ml volume

Samples >1ml and less than 5ml are filled up using buffer *SU* (*Add-On 10*) Transfer the sample by pipetting into a *50ml Centrifuge Tube* (*Add-On 10*). Then add buffer *SU* using a disposable 5ml pipette or pipette tip until reaching the 5ml mark of the tube. Discard pipette/pipette tip with excess buffer *SU*. Continue with part B.

B) Sample pre-treatment and DNA Isolation

Per sample:

- 1. Pipette **5ml sample** or **filled-up sample** (see part A) into a *50ml Centrifuge Tube*.
 - Add **2ml buffer** *CM* (Kit 1), vortex for 15s.
 - Let stand at room temperature (+18 to +25 $^{\circ}$ C) for 5min.
- Add 2ml buffer DB1 (Kit 1).
 Add 10µl MolDNase B (Kit 2), vortex for 15s.
 Incubate at room temperature (+18 to +25°C) for 15min.
- 3. Centrifuge at 9,500xg, 10min. Carefully decant supernatant.
- 4. Resuspend pellet in **1ml buffer RS** (Kit 1). Transfer by pipetting into a *ST tube* (Kit 1).

Continue with Short Manual of *UMD-SelectNA™* (page 5, step 5).

Add-On 10 / DNA Isolation* Part 2: Add-on kit for *UMD-SelectNA™*



Protocol 2:

Large Size Sample DNA Isolation (>5 to 10ml Body Liquids)

To be used with **Kit 1** and **Kit 2** of **UMD-SelectNA**TM:

Kit 1 and Add-On 10 Kit: Extraction Buffers (+18 to +25°C):

Unpack buffer bottles: SU - CM - DB1 (Add-On 10) - RS - RL (Kit 1)

Kit 2: Enzymes & Reagents (-15 to -25°C):

A) Fill up procedure for samples less than 10ml volume

Samples >5ml and less than 10ml are filled up using buffer *SU* (*Add-On 10*). Transfer the sample by pipetting into a *50ml Centrifuge Tube* (*Add-On 10*). Then add buffer *SU* using a disposable 5ml pipette or pipette tip until reaching the 10ml mark of the tube. Discard pipette/pipette tip with excess buffer *SU*. Continue with part B.

B) Sample pre-treatment and DNA Isolation

Per sample:

- 1. Pipette **10ml sample** or **filled-up sample** (see part A) into a *50ml Centrifuge Tube*.
 - Add **4ml buffer** *CM* (Kit 1), vortex for 15s.
 - Let stand at room temperature (+18 to +25°C) for 5min.
- Add 4ml buffer DB1 (Kit 1).
 Add 10μl MolDNase B (Kit 2), vortex for 15s.
 Incubate at room temperature (+18 to +25°C) for 15min.
- 3. Centrifuge at 9,500xg, 10min. Carefully decant supernatant.
- 4. Resuspend pellet in **1ml buffer RS** (Kit 1). Transfer by pipetting into a *ST tube* (Kit 1).

Continue with Short Manual of *UMD-SelectNA™* (page 5, step 5).