

GeneProof™ Cytomegalovirus (CMV) PCR Kit



1. LIST OF PRODUCT VARIANTS

Product	Package	REF
GeneProof™ Cytomegalovirus (CMV) PCR Kit	25 reactions	CMV/GP/025
GeneProof™ Cytomegalovirus (CMV) PCR Kit	100 reactions	CMV/GP/100

2. INTENDED PURPOSE AND USE

The intended purpose of the device is related to the medical context and clinical conditions under which the device may be used.

Clinical Conditions	CMV is implicated in infectious mononucleosis; CMV implicated diseases can present in many ways including fever, pneumonia, pneumonitis, colitis, hepatitis, myocarditis, esophagitis, gastrointestinal ulcerations, diarrhoea, retinitis, visual impairment, blindness, polyradiculopathy, transverse myelitis, encephalopathy, encephalitis, seizures, and coma; congenital CMV infections can result in non-specific symptoms and include rhinitis, pharyngitis, myalgia, arthralgia, headache, and fatigue
Indication	<i>In vitro</i> diagnostic medical device
Regulatory Status	CE ₂₇₉₇ IVD / Regulation (EU) 2017/746
Summary of Safety and Performance	See EUDAMED – European Database on Medical Devices at ec.europa.eu
Function	Diagnostics, aid to diagnosis and for monitoring
Specific Information	Testing of immunocompromised patients
What is Detected / Target	Cytomegalovirus (CMV; Human beta herpesvirus 5)
Automated / Manual Detection	Manual
Type of Analysis	Qualitative and quantitative
Validated Specimen	DNA extracted from CSF, plasma, serum, urine, whole blood (EDTA)
Testing Population	EU population
Intended User	For professional use in laboratories with trained staff
Test Principle	Real-time polymerase chain reaction (PCR) – amplification of the specific Target Sequence and detection using TaqMan probes with fluorophore-based detection

3. TECHNICAL SPECIFICATIONS

Target Sequence	Gene encoding the IE1 protein						
Analytical Specificity	Human Cytomegalovirus (CMV), 100 %						
Analytical Sensitivity (LoD with 95% probability)	Sample Processing	CSF	Plasma	Serum	Urine	Whole blood	
	GeneProof™ PathogenFree DNA Isolation Kit	134.28 IU/ml	122.59 IU/ml	87.52 IU/ml	255.48 IU/ml	172.44 IU/ml	
	croBEE 201A Nucleic Acid Extraction Kit	411.43 IU/ml	165.24 IU/ml	228.15 IU/ml	745.68 IU/ml	117.70 IU/ml	
	myCROBE/croBEE 2.0 Universal Extraction Kit	630.34 IU/ml	281.37 IU/ml	82.49 IU/ml	287.95 IU/ml	431.21 IU/ml	
Diagnostic Specificity	94.21 % (CI _{95%} : 88.01 % - 97.44 %)						
Diagnostic Sensitivity	98.32 % (CI _{95%} : 95.90 % - 99.38 %)						
Positive Predictive Value	97.67 % (CI _{95%} : 95.04 % - 98.97 %)						
Negative Predictive Value	95.80 % (CI _{95%} : 89.98 % - 98.44 %)						
Linear Range [IU/ml]	Extraction Method	with precision of ± 0.5 log	CSF	Plasma	Serum	Urine	Whole blood
	GeneProof™ PathogenFree DNA Isolation Kit		10 ¹⁰ – 10 ^{2.5}	10 ¹⁰ – 10 ^{2.5}	10 ¹⁰ – 10 ²	10 ¹⁰ – 10 ³	10 ¹⁰ – 10 ^{2.5}
	croBEE 201A Nucleic Acid Extraction Kit		10 ¹⁰ – 10 ³		10 ¹⁰ – 10 ^{2.5}	10 ¹⁰ – 10 ³	10 ¹⁰ – 10 ³
	myCROBE/croBEE 2.0 Universal Extraction Kit		10 ¹⁰ – 10 ³	10 ¹⁰ – 10 ^{2.5}	10 ¹⁰ – 10 ^{2.5}	10 ¹⁰ – 10 ^{2.5}	
Dynamic Range	10 ¹⁰ IU/ml – LoD (LoD varying according to the extraction and material used)						
Trueness (of expected concentration)	Extraction Method	CSF	Plasma	Serum	Urine	Whole blood	
		CI _{95%}	CI _{95%}	CI _{95%}	CI _{95%}	CI _{95%}	
	GeneProof™ PathogenFree DNA Isolation Kit	-0.06 log -0.12 – -0.01 log	-0.03 log -0.12 – -0.05 log	0.10 log -0.05 – -0.24 log	-0.15 log -0.28 – -0.02 log	-0.09 log -0.16 – -0.01 log	
	croBEE 201A Nucleic Acid Extraction Kit	-0.06 log -0.13 – -0.02 log	0.06 log -0.06 – -0.18 log	0.06 log -0.08 – -0.20 log	-0.14 log -0.26 – -0.02 log	-0.06 log -0.14 – -0.02 log	
myCROBE/croBEE 2.0 Universal Extraction Kit	-0.09 log -0.17 – -0.00 log	-0.04 log -0.11 – -0.02 log	0.06 log -0.08 – -0.19 log	-0.11 log -0.22 – -0.01 log	-0.10 log -0.19 – -0.01 log		
Precision - repeatability	• Intra-assay SD of log concentration = 0.054 (CI _{95%} : 0.043 – 0.070)						
Precision - reproducibility	• Inter-assay SD of log concentration = 0.059 (CI _{95%} : 0.038 – 0.130)						
	• Inter-lot SD of log concentration = 0.062 (CI _{95%} : 0.040 – 0.136)						
	• Total SD of log concentration = 0.062 (CI _{95%} : 0.040 – 0.137)						
Reporting Units	IU/ml						
Conversion Factor	1 IU = 1 cp						

Metrological Traceability	CMV NIBSC 09/162 (1 st WHO International Standard for Human Cytomegalovirus for Nucleic Acid Amplification Techniques)		
Extraction / Inhibition Control	PCR inhibition and DNA extraction efficiency control by the Internal Control (IC)		
Validated Extraction Methods	croBEE 201A Nucleic Acid Extraction Kit myCROBE/croBEE 2.0 Universal Extraction Kit GeneProof™ PathogenFree DNA Isolation Kit		
Applied Instruments	Instrument Name	CMV	Internal Control (IC)
	croBEE Real-Time PCR System	FAM	HEX
	AMPLilab Real-Time PCR System	FAM	HEX
	Applied Biosystems 7300 / 7500 Real-Time PCR System	FAM	JOE
	AriaMx Real-Time PCR System	FAM	HEX
	BioQuant-96 Real-Time PCR System	FAM	HEX
	CFX Connect™ / CFX96™/ Dx Real-Time PCR Detection System	FAM	HEX
	CFX Opus 96 Real-Time PCR System	FAM	HEX
	Gentier 96E/96R Real-Time PCR System	FAM	HEX
	LightCycler® 480	FAM	HEX
	LineGene 9600 / 9600 Plus	FAM	HEX
	Mic qPCR Cyclor	FAM	HEX
	Montania 4896 Real-Time PCR termocykler	FAM	HEX
	QuantStudio™ 3 / 5 Real-Time PCR System	FAM	VIC
Rotor-Gene 3000 / 6000 / Q	FAM	JOE	
SLAN® Real-Time PCR System	FAM	HEX	
StepOne™ / StepOne Plus™ Real-Time PCR System	FAM	HEX	
Detection Channels	FAM (CMV), HEX/JOE/VIC (IC)		
External Quality Assessment	Regularly tested using QCMD and INSTAND e.V. External Quality Assessment Panels - results at www.geneproof.com		

4. INTERFERENCES

The evaluation and setting of pathological values for interference testing was performed according to CLSI guidelines EP7-A2 and guidelines and recommendations of Czech Society of Clinical Biochemistry.

Endogenous and Exogenous Interferences

Tested Substance	Tested Level(s)	Observed Interference	Tested Substance	Tested Level(s)	Observed Interference
PLASMA					
Albumin	60 g/l	Partial	Haemoglobin	2 g/l	None
Bilirubin	342 µmol/l	Partial	Urea	42.9 mmol/l	None
Glucose	55 mmol/l	None	Uric acid	1.4 mmol/l	None
Caffeine	308 µmol/l	None	Prednisone	0.84 µmol/l	Partial
Ibuprofen	2425 µmol/l	Partial	Vancomycin	69 µmol/l	None
Fluconazole	245 µmol/l	Partial	Valganciclovir	20 mg/l	Partial
Citrate	19 g/l	None	-	-	-
SERUM					
Albumin	60 g/l	Partial	Haemoglobin	2 g/l	None
Bilirubin	342 µmol/l	None	Urea	42.9 mmol/l	None
Glucose	55 mmol/l	None	Uric acid	1.4 mmol/l	None
URINE					
Albumin	5 % (w/v)	None	pH	Basic condition (pH 9)	Partial
Bilirubin	1 % (w/v)	Partial	Urea	600 mmol/l	Partial
Glucose	0.1 % (w/v); 1 % (w/v)	Partial	Uric acid	5 mmol/l	Partial
pH	Acidic condition (pH 4)	Partial	-	-	-
CSF					
Glucose	55 mmol/l	None	Albumin	60 g/l	None
Lactic acid	16.5 mmol/l	None	-	-	-
WHOLE BLOOD					
EDTA	2.2 g/l	None	Heparin	30 IU/ml	None

NOTE: In the case of partial interference, inhibition may occur with the risk of a false negative result at a given concentration of interferent.

5. KIT CONTENTS

Reagent	Content	Vial Title	Cap Colour	Guaranteed Volume [µl]	Number of Vials	
					CMV/GP/025 – 25 rxn	CMV/GP/100 – 100 rxn
Master Mix	Mixture of PCR enzymes, target specific primers and TaqMan probes in buffer	MasterMix CMV	Blue	750	1	4
Calibrator	DNA oligonucleotide in buffer	Calibrator A CMV 10 ⁴ IU/µl	Black	200	1	1
		Calibrator B CMV 10 ³ IU/µl	Brown	200	1	1
		Calibrator C CMV 10 ² IU/µl	White	200	1	1
		Calibrator D CMV 10 ¹ IU/µl	Transparent	200	1	1
Internal control	Plasmid DNA in buffer	Internal Control CMV	Red	1000	1	2

DESCRIPTION OF REAGENTS AND LIMITATIONS

Mixtures in this product (Master Mix, Calibrators and Internal Control) are not classified as dangerous according to Regulation (EC) No 1272/2008.

6. CALIBRATOR INFORMATION

Use of all 4 calibrators is necessary for a correct sample quantification. An automatic quantification based on the analysis of calibrators is generated automatically as a part of an analytical process performed in the PCR instrument. Each calibrator consists of target specific DNA. Each calibrator must be designated as „standard“ in the instrument (thermocycler). The concentration of each calibrator must be entered in the data analysis software during PCR plate set up.

NOTE: For qualitative detection, the Calibrator C 10² IU/µl serves as a positive control.

7. TRANSPORT AND STORAGE

Storage Conditions	(-20 ± 5) °C
Transport Conditions	-20 °C and below
In-use Stability	Thaw a maximum of 5 times or use within 30 days after the first use of a particular vial, whichever comes first

8. ASSAY PROCEDURE

SPECIMEN COLLECTION, TRANSPORTATION AND HANDLING

Samples for DNA extraction must be collected and transported following professional guidelines.

Samples for DNA extraction must be transported and processed by the laboratory as soon as possible.

CSF specimens should be transported at (5 ± 3) °C. If specimens cannot be processed immediately, CSF should be stored at -20 °C or at -70 °C or lower.

Plasma specimens should be collected in EDTA-containing collection tubes and transported at (5 ± 3) °C. Plasma specimens can be stored at (5 ± 3) °C for up to 7 days from blood collection.

Serum specimens should be transported at (5 ± 3) °C and can be stored at (5 ± 3) °C for up to 4 days from collection.

Urine specimens should be collected as first catch of 10 to 20 ml in a clean collection cup and refrigerated immediately at (5 ± 3) °C. Urine specimens can be stored at (5 ± 3) °C for up to 4 days.

Whole blood specimens should be collected in EDTA-containing collection tubes and transported at room temperature within 24 hours. Whole blood specimens can be stored at 4 °C for up to 14 days from collection.

NOTE:

For more information see Instructions for Use of the corresponding extraction kit.

NUCLEIC ACID PURIFICATION

1. Prepare specimens for the assay according to the corresponding extraction kit manual.
2. Thaw required number of Internal Control (IC or UNIC*) vials, mix and briefly centrifuge.
3. Add the Internal Control (IC or UNIC) directly to the sample at the beginning of the extraction process so that 1 µl of the final elution volume contains 0.1 µl of the IC:

Elution Volume	25 µl	50 µl	100 µl	200 µl
Internal Control (IC or UNIC)	2.5 µl	5 µl	10 µl	20 µl

4. Continue extraction according to the appropriate protocol.

NOTE:

*If using *UNIC = GeneProof™ Universal Internal Control (more information in chapter 12. Additional Products), see Instructions for Use of GeneProof™ Universal Internal Control.*

The samples of DNA extracted from CSF, plasma, serum, urine and whole blood can be stored for 7 days at (5 ± 3) °C, at (20 ± 5) °C or at (-20 ± 5) °C. The samples can be frozen and thawed 3 times after the extraction.

PCR SETUP PROTOCOL

5. Thaw required vials and reagents completely.
6. Vortex gently and centrifuge briefly all vials before setting up the PCR run.

NOTE:

Keep the reagents at (5 ± 3) °C for the shortest time possible until the PCR reaction is set up.

7. Add 30 µl of Master Mix into PCR tubes.
8. Add 10 µl of the extracted nucleic acid sample or 10 µl of Calibrator into each individual PCR tube and mix by pipetting. The total reaction mix volume is 40 µl.
9. Close the tubes, centrifuge shortly, insert them into the real-time PCR device and amplify according to the following PCR profile.

NOTE:

It is recommended to include at least 1 negative control and at least 1 complete set of calibrators (for a quantitative kit) in each individual PCR run. Use nuclease-free water as your own negative control (not provided). For more information see chapter 10. Run Validity.

AMPLIFICATION PROFILE

Follow the thermocycler manufacturer's manual for setting the instrument and for analysis.

Universal PCR Profile

Step	Process	Temperature [°C]	Time	Cycles	Fluorescence Acquisition
1	UNG decontamination/ Reverse Transcription	42	15 min	1 cycle	
2	Initial denaturation	95	10 min	1 cycle	
3	Denaturation	95	5 s	45 cycles	FAM, HEX/JOE/VIC
	Annealing	60	40 s		
	Extension	72	20 s		

9. INTERPRETATION OF RESULTS

Channel FAM (CMV)	Channel HEX/JOE/VIC (IC)	Result	Interpretation
+	+	Valid	CMV positive
+	-	Valid	CMV positive
-	+	Valid	CMV negative
-	-	Invalid	-

NOTE: For evaluation of PCR run see chapter 10. Run Validity.

10. RUN VALIDITY

OVERALL VALIDITY OF DETECTION

	Signal	Channel	Run Validity	Recommendation
Calibrator C 10 ² (qualitative detection) Or Calibrator Set (quantitative detection)	+	FAM	Valid	-
Calibrator C10 ² (qualitative detection) or Calibrator Set (quantitative detection)	-	FAM	Invalid	Repeat PCR run
Negative control	-	FAM	Valid	-
Negative control	+	FAM	Invalid	Repeat PCR run

NOTE: If the issue persists, please contact Customer Support, see Contact information.

11. QUANTITATIVE DETECTION EVALUATION

Use the following formula to calculate the viral load concentration in IU/ml for manual extraction (using GeneProof™ PathogenFree DNA Isolation Kit):

VLC – Viral load concentration [IU/ml]

SC - Sample concentration [IU/µl]

EV - Elution volume [µl]

IV - Isolation volume [µl]

$$VLC = \frac{SC \times EV \times 10^3}{IV}$$

To easily calculate pathogen concentrations, use the calculator at www.geneproof.com, where manual or automated nucleic acid extraction used is considered.

VALIDITY OF QUANTITATIVE DETECTION

Channel	Calibrators				Result	Recommendation
	A 10 ⁴	B 10 ³	C 10 ²	D 10 ¹		
Target-specific channel (FAM)	++++	+++	++	+	Valid accurate quantification	-
Internal Control channel (HEX/JOE/VIC)	+/-	+/-	+/-	+/-		
R ²	≥0.98					
Target-specific channel (FAM)	++++	+++	++	+	Reduced quantification accuracy	Repeat PCR run
Internal Control channel (HEX/JOE/VIC)	+/-	+/-	+/-	+/-		
R ²	<0.98					
Target-specific channel (FAM)	No signal of one or more calibrators				Invalid quantification	Repeat PCR run
Internal Control channel (HEX/JOE/VIC)	+/-	+/-	+/-	+/-		
R ²	N/A					

R² – Determination coefficient – parameter evaluating the quality of standard curve

NOTE: If the issue persists, please contact Customer Support.

12. ADDITIONAL PRODUCTS

GeneProof™ Universal Internal Control

GeneProof™ Universal Internal Control (UNIC) is intended to be used as an internal control for all microbiological GeneProof PCR kits and as an alternative product to Internal Controls included in all GeneProof microbiological PCR kits. The UNIC works only in combination with the GeneProof PCR kits. It is intended to simplify the user's workflow where multiple detection kits are used with a single nucleic acid extract. For more details see the Instructions for Use for UNIC.

Product	REF
GeneProof™ Universal Internal Control	UNIC/GP/050

NOTE: IC is applied to the sample only once. Add UNIC instead of IC from the package of the PCR kit. Do not add both IC and UNIC to the same sample at the same time.

13. MATERIALS AND DEVICES REQUIRED BUT NOT PROVIDED

CONSUMABLE MATERIALS

96-well reaction plates or PCR tubes (0.2 ml volume), pipette tips with filters, powder-free gloves, biohazard waste bin, nuclease-free water

DEVICES

Real-time PCR instrument (see chapter 3. Technical Specifications), nucleic acid extraction system or kit (see chapter 3. Technical Specifications), desktop centrifuge (for 0.2 ml PCR tubes or 96-well plates), vortex mixer, freezer (-20 ± 5) °C, refrigerator (5 ± 3) °C, cooling rack

14. WARNINGS, PRECAUTIONS AND PROCEDURE LIMITATIONS

- Patient management decisions should never be based solely on the results from this test. Other laboratory and clinical factors must also be considered in making clinical decisions.
- Any serious incident that occurred in relation to using of the GeneProof PCR Kit shall be reported to the manufacturer and to the competent local authority.
- Use separate working places for sample preparation / nucleic acid extraction and amplification reactions. Never introduce an amplified product in reagent and/or nucleic acid extraction (sample preparation) area.
- Dispose of unused reagents and waste in accordance with national, federal, state or local regulations.
- Use all necessary protective equipment (protective disposable gloves, a laboratory coat and eye protection) when handling specimens and kit reagents.
- Avoid microbial and ribonuclease contamination of the reagents when removing aliquots from reagent vials.
- Use RNase- and DNase-free filter pipette tips only.
- Use new tips for each pipetting step.
- Close the kit components vials immediately after use and never interchange lids.
- Do not pool reagents from different lots or from different vials within the same lot.
- Do not substitute the reagents between different lots.
- Do not use reagents from damaged or leaking vials.
- Be very careful when handling the Calibrators or the clinical material; incorrect handling could result in contamination and the consequent defectiveness of the kit components! The manufacturer is not responsible for the kit defectiveness due to incorrect handling.











Procedure Limitations:

- Read the whole Instructions for Use properly before starting the manipulation. Not following these instructions can lead to an erroneous result which can cause misdiagnosis or inappropriate treatment!
- Appropriate specimen collection, transport, storage and processing procedures are required for the optimal performance of this test.
- Do not use the kit after its expiry date.
- The presence of UNG decontamination step reduces the risk of amplicon contamination. However, contamination from very high levels of amplicons can be avoided only by good laboratory practices and careful adherence to the procedures specified in these Instructions for Use. All reagents should be closely monitored for impurity and contamination. Any suspicious reagents should be discarded.
- This product is designed for use with the applied PCR instruments and validated extraction methods mentioned in chapter 3. Technical Specifications.
- Proper homogenization of used clinical material is necessary for quantitative analysis of CMV DNA. Homogenization by a short vortex and short spin is recommended.

Clinical Limitations:

- Though rare, mutations within the highly conserved regions of the viral genome covered by GeneProof™ Cytomegalovirus (CMV) PCR kit primers and/or probes may result in the under-quantitation of or failure to detect the virus.
- Detection of CMV DNA is dependent on the number of virus particles present in the specimen and may be affected by specimen collection methods and patient factors.
- Use only with validated specimens (see chapter 2. Intended Purpose and Use) otherwise incorrect results could occur.

15. EXPLANATION OF SYMBOLS

Symbol	Explanation	Symbol	Explanation
	This product complies with the relevant EU requirements		Lot number
	<i>In vitro</i> diagnostic medical device		Contains sufficient amount for n-tests
	Catalogue number		Temperature limitation
	Manufacturer		Expiry date
	Read electronic Instructions for Use		Unique Device Identifier

www.geneproof.com/ifu

16. REFERENCES

- Institute, C.L.S., *Collection, Transport, Preparation, and Storage of Specimens for Molecular Methods*, in 2nd ed. *CLSI guideline MM13*. 2020. p. 94.
- Abdul-Ali, D., et al., *Cytomegalovirus DNA stability in EDTA anti-coagulated whole blood and plasma samples*. *J Clin Virol*, 2011. **52**(3): p. 222-4.
- Xie, L., et al., *Effects of storage time on Cytomegalovirus DNA stability in plasma determined by quantitative real-time PCR*. *J Virol Methods*, 2014. **207**: p. 196-9.
- Baleriola, C., et al., *Stability of hepatitis C virus, HIV, and hepatitis B virus nucleic acids in plasma samples after long-term storage at -20 degrees C and -70 degrees C*. *J Clin Microbiol*, 2011. **49**(9): p. 3163-7.
- Satsangi, J., et al., *Effect of heparin on polymerase chain reaction*. *Lancet*, 1994. **343**(8911): p. 1509-10.
- Holodniy, M., et al., *Inhibition of human immunodeficiency virus gene amplification by heparin*. *J Clin Microbiol*, 1991. **29**(4): p. 676-9.
- Beutler, E., T. Gelbart, and W. Kuhl, *Interference of heparin with the polymerase chain reaction*. *Biotechniques*, 1990. **9**(2): p. 166.
- Bai, X., et al., *Heparin interference with reverse transcriptase polymerase chain reaction of RNA extracted from lungs after ischemia-reperfusion*. *Transpl Int*, 2000. **13**(2): p. 146-50.
- Black, C.M., *Current methods of laboratory diagnosis of Chlamydia trachomatis infections*. *Clin Microbiol Rev*, 1997. **10**(1): p. 160-84.
- Akinlade, F. and S. Whiting. *Pathology First Biochemistry Reference Intervals*. November 2017; Version 1.3: Available from: http://www.southend.nhs.uk/media/180421/pf_biochemistry_reference_intervals.pdf.

17. MODIFICATIONS INTRODUCED IN THE LATEST VERSION

In connection with implementation of the *In vitro* Diagnostic Medical Devices Regulation (EU) 2017/746, a new version of the Instructions for Use has been issued, which meets all requirements of this legislation.

Customer Support

Tel.: +420 730 176 222
E-mail: support@geneproof.com

Orders

Tel.: +420 543 211 679
E-mail: sales@geneproof.com

 GeneProof a.s.
Václavská 101/119 /Dolní Heršpice/ CZ-619 00 Brno / +420 543 211 679
info@geneproof.com

